

## Invited Review

# Passive Protection Against Diarrheal Disease

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Diarrheal disease is a major health problem causing high mortality and morbidity in developing countries. In developed countries acute diarrheal disease is an important cause of hospitalization. Rotavirus is the major cause of infectious diarrhea in infants and young children globally (1). Other important enteric pathogens, particularly in developing countries, include enterotoxigenic and enteropathogenic *Escherichia coli*, *Shigella*, *Campylobacter* species, *Vibrio cholerae*, and *Cryptosporidium* (2).

Vaccines to immunize actively against these pathogens have reached various stages: licensure, controlled trials of efficacy, and clinical trials of safety and immunogenicity (3). Another approach to protection against diarrheal disease has been the use of passively acquired antibodies from either serum or colostrum. This review concentrates on studies in which bovine immunoglobulins, either in colostrum or as a bovine immunoglobulin concentrate, have been used to prevent or treat diarrheal disease.

There are a number of settings in which passive protection may be valuable to prevent diarrheal illness, particularly in the absence of active immunization. Rotavirus infection is responsible for nosocomial infection in ~20% of infants in the United States (3) and Australia (4). Passive protection could possibly lessen this high incidence of nosocomial rotavirus diarrhea (5,6). Specific settings in which passive protection may be of value in preventing infection include neonatal nurseries (7) and daycare centers, within families (8), and in immunocompromised people (9,10).

For many GI infections the most important pro-

TECTIVE factor is the presence of specific antibody in the lumen of the small intestine. This fact has certainly been proven for rotavirus infection (11), for which the antibody in humans is secretory IgA. Human milk has been shown to contain specific secretory IgA antibodies to many enteropathogens (12). For example, colostrum and breast milk have been shown to contain antibodies to the four major human rotavirus serotypes (13). Breastfed infants, however, still develop rotavirus diarrhea (14,15), which may be due to an overwhelming antigen load or to the absence of antibodies against the infecting serotype.

Many of the studies involving passive protection have been carried out using bovine colostrum or bovine milk immunoglobulin concentrates. The colostrum product was taken from nonimmunized cows or from cows hyperimmunized against various pathogens (5-34). Some studies have used human  $\gamma$ -globulin or human serum immunoglobulin (7,9,10). In ruminants, the predominant milk antibody is IgG<sub>1</sub>, which is mainly serum-derived and has specificity for enteric pathogens (34). IgG<sub>1</sub> has been shown passively to protect against infection by various enteric pathogens in animals and humans (Table 1). This finding lends support to the concept that ruminant IgG<sub>1</sub> may replace secretory IgA in providing lactogenic immunity. Saif (40) reviewed the evidence supporting similarities and differences between ruminant IgG<sub>1</sub> and secretory IgA, which includes resistance to proteolytic enzymes; the predominance of IgG<sub>1</sub> in milk and its specificity against enteric viruses; and the increase in IgG<sub>1</sub> but not IgA milk antibodies after intestinal administration of antigens in pregnant cows. In humans, bovine immunoglobulin IgG<sub>1</sub> has been shown to survive transit through the gut in colostrum (46), although there are conflicting reports regarding the survival of human immunoglobulin and bovine immunoglobulin concentrates (10,24,47).

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TABLE 1. Human pathogens studied using passive protection

Enteric pathogens (and references)	Study subjects
<i>Campylobacter jejuni</i> (35)	Calves
<i>Clostridium difficile</i> (36)	Hamsters
<i>Cryptosporidium parvum</i> (19,22,23,26-29,32,34,37,38)	Humans and mice
Enteropathogenic <i>E. coli</i> (17,30)	Humans
Enterotoxigenic <i>E. coli</i> (25)	Humans
Rotavirus (5-11,18,20,21,30,39-43)	Humans, lambs, calves, and pigs
<i>Shigella flexneri</i> 2a (33)	Humans
<i>Vibrio cholerae</i> (24,44,45)	Humans and rabbits

Losonsky et al. (10) showed that human serum immunoglobulin survived passage through the gut in immunocompromised children in an immunologically active form. The studies of Hilpert et al. (21) and McClead et al. (24) demonstrated partial recovery of functionally active bovine immunoglobulin preparations against rotavirus and cholera, respectively, following passage through the gut. Using an in vitro model of gut digestion, Petschow and Talbot (47) demonstrated a significant reduction in rotavirus-neutralizing titer in a bovine colostrum immunoglobulin concentrate by gastric acid, pepsin, and selected pancreatic enzymes. While comparative studies have not been carried out, it is possible that bovine IgG<sub>1</sub> survives passage through the gut more completely when given as a colostrum product rather than as an immunoglobulin concentrate. The work of Ebina et al. (31) would support this concept, as they found whole colostrum but not specific immunoglobulins to be protective.

The level of antibody present in colostrum and breast milk can be markedly increased by parenteral vaccination of the pregnant cow prior to delivery of the calf. While immunization of human mothers during pregnancy or immediately postpartum to boost rotavirus antibodies in breast milk has been considered, there are no published results of such studies (48). Brussow et al. (49) found that cows have elevated preimmunization titers in serum and colostrum to the four major human rotavirus serotypes (49), but this has not been a universal finding (50). Heterologous immunity is probably related to natural exposure to bovine rotavirus serotypes resembling human rotaviruses. Cows immunized with a single rotavirus serotype (SA11 serotype 3) produced an increase in neutralizing antibodies not only to serotype 3 but also serotypes 1, 2, 4, and 6. Similarly, cows immunized with SA11 produced co-

lostral antibodies that protected piglets against challenge with two porcine rotavirus serotypes (42). Brussow et al. (49) also noted the induction of cross-neutralization antibodies by a single-serotype vaccination of cows with rotavirus. They suggest this response may be due to the recognition by the bovine immune system of a minor neutralizing antigen common to different rotavirus serotypes. There is also evidence that human rotaviruses of different serotypes may share immunodominant neutralization sites (51).

Heterotypic responses have also been noted in adult human volunteers following oral vaccination with a single-serotype rotavirus vaccine candidate (52). Young infants do not produce a heterotypic response, probably because of lack of prior exposure to rotavirus (52). The heterotypic response of the bovine immune system to a single rotavirus strain simplifies the commercial production of hyperimmune colostrum. The common human rotavirus serotypes are difficult to propagate, whereas SA11 is easily cultivated and can stimulate high-titer rotavirus antibodies that neutralize the four major human rotavirus serotypes (49).

## PASSIVE PROTECTION STUDIES IN HUMANS

### Rotavirus

Barnes et al. (7) were the first to show that oral immunoglobulin containing rotavirus antibody could modify the course of rotavirus infection. Low-birth-weight infants in a special-care nursery fed human  $\gamma$ -globulin, 5 ml four times daily, had a reduced incidence of rotavirus acquisition and reduced severity of rotavirus diarrhea.

Using hyperimmune bovine colostrum from cows immunized with human rotavirus Wa strain (serotype 1), Ebina et al. (18) were able to protect infants in an orphanage from acquiring rotavirus infection during a rotavirus outbreak. Infants fed commercial milk were not protected. Importantly, two of the infants fed rota-colostrum who remained asymptomatic showed rises in complement-fixing antibody. Thus, the rota-colostrum did not prevent immunological responses to natural rotavirus infection.

More recently, Ebina et al. (31) confirmed their earlier work in an animal model and in infants. They also showed that purified IgG, IgM, and IgA from rota-colostrum were not protective, possibly due to inactivation by GI proteolytic enzymes—which did not occur using the rota-colostrum. Petschow and

Talbot (47) showed that both acid and trypsin significantly reduce the biological activity of bovine milk immunoglobulins directed against rotavirus.

Some studies have provided evidence for the role of hyperimmune bovine colostrum in protecting against rotavirus cross-infection (5,6). Davidson et al. (5) fed hospitalized children aged 3–15 months with bovine colostrum containing high-titer antibodies to the four major human rotavirus serotypes. Nine of 65 control infants, but no treated infant, acquired symptomatic rotavirus infection during the treatment period. When diarrhea occurred, parents of children with symptomatic rotavirus infection were seven times more likely to seek medical attention than were parents of children with nonrotavirus diarrhea. A similar study was carried out in Hong Kong and India (6), confirming the earlier findings. In this study, the control group was fed colostrum containing no antibodies to rotavirus. No child in the treatment group developed symptomatic rotavirus infection versus 13 in the control group ( $p < 0.01$ ). This result highlights that it was the presence of rotavirus antibody in colostrum rather than colostrum itself that provided protection against rotavirus infection. It is likely that the antibody titer is important, with higher antibody levels allowing for some intraluminal degradation of product while maintaining sufficient protective activity (43). The latter study (6) also showed that hyperimmune colostrum could protect against more than one rotavirus serotype.

The approaches discussed so far used the hyperimmune colostrum as a food additive. Several recent studies have used infant formula supplemented with hyperimmune bovine immunoglobulin, with conflicting results (8,30). A study by Brunser et al. (30) in Chile using formula containing anti-rotavirus antibodies and anti-enteropathogenic *E. coli* antibodies was unsuccessful, most likely related to the low level of antibody (43). A trial from Charleston, SC, U.S.A. (8), was successful in reducing symptomatic rotavirus infection in healthy infants in the community but had no effect on the actual incidence of rotavirus infection. This failure to prevent infection while reducing symptoms almost certainly relates to the level of antibody given. Schaller et al. (43) showed in a gnotobiotic piglet model that at very high levels of antibody administration, both symptoms and rotavirus excretion could be abolished. However, it may not be economically feasible to use this approach in infant formula. Similarly, while the addition of colostrum antibodies to infant

formula may be helpful in young infants, it does not provide protection once infants are weaned, which usually happens around 12 months of age when infants are still at risk of severe rotavirus illness. The use of a food supplement would be more practical for older infants.

Several studies have reported on the efficacy of human milk or human serum immunoglobulin in managing the immunocompromised child with chronic rotavirus infection (9,10). Other experimental approaches to the provision of passive prophylaxis have used egg yolk immunoglobulin (53,54), human milk (55), and human intestinal mucins (56), but these approaches have yet to be tested in humans. Saavedra et al. (57) found that infant formula supplemented with *Bifidobacterium bifidum* and *Streptococcus thermophilus* can reduce the incidence of acute diarrhea and rotavirus shedding in infants admitted to the hospital.

Hyperimmune bovine colostrum concentrate (21) and oral human serum immunoglobulin have also been used in the treatment of acute and chronic rotavirus diarrhea (59,60). Mitra et al. (58) have recently shown therapeutic efficacy of hyperimmune bovine colostrum in the treatment of rotavirus gastroenteritis in children aged six to 24 months. The treatment group received 100 ml of hyperimmune colostrum three times daily and showed a significant reduction in duration and severity of the diarrhea. Hilpert et al. (21) fed hyperimmune bovine immunoglobulin concentrate in a daily dose of 2 g/kg body weight for 5 days to 75 infants hospitalized with acute rotavirus gastroenteritis. A 10-kg child thus received 20 g/day of bovine immunoglobulin; no side effects were seen. Hilpert et al. noted a decrease in duration of rotavirus excretion but no effect on clinical symptoms. More recently, Guarino et al. (58) fed a single dose of human serum immunoglobulin (300 mg/kg) to hospitalized children with acute rotavirus diarrhea in a double-blind placebo-controlled trial. They found a significant decrease in the duration of diarrhea, viral excretion, and hospital stay in the treated group. The cost of treatment was high, at \$200 per 10-kg infant, but this expense may be offset by the decreased length of hospital stay.

#### *Escherichia coli*

Lyophilized immunoglobulins obtained from colostrum of cows immunized with several enterotoxigenic *E. coli* serotypes, fimbria types, *E. coli* heat-labile enterotoxin, and cholera toxin were shown to

provide complete protection against enterotoxigenic *E. coli* infection in 10 adult volunteers (25). This approach shows that bovine milk immunoglobulins may have a role in the prevention of travelers' diarrhea.

Brunser et al. (30), however, were unable to show any protective benefit of supplementing infant formula with bovine milk immunoglobulin concentrate against the major enteropathogenic *E. coli* serogroups in a small-scale field trial in Chile. This failure probably relates to the low level of antibody in the formula.

Sixty children aged 10 days to 18 months with diarrhea due to enteropathogenic *E. coli* were treated daily for 10 days with 1 g/kg hyperimmune anti-enteropathogenic *E. coli* immunoglobulin (17). The treatment was effective in eliminating the *E. coli* in 43 of 51 children infected with strains present in the vaccine but in only one of nine infected with strains not incorporated in the vaccine. These results suggest that specific milk immunoglobulin may be an effective treatment for *E. coli* infections.

#### *Cryptosporidium*

A number of small studies using either hyperimmune bovine colostrum containing cryptosporidial antibodies (19,23,27,29,34) or nonspecific bovine immunoglobulin concentrate (32) or colostrum (22,26) have been reported with variable but encouraging results. Most patients were immunosuppressed due to HIV infection, congenital hypogammaglobulinemia, or leukemia. The failures in this approach occurred mainly in patients given nonspecific bovine immunoglobulin or colostrum, again highlighting the importance of specific antibody. At present, there is no specific therapy for *Cryptosporidium*, but the results of the reported studies suggest that further double-blind controlled studies of a hyperimmune anticryptosporidial product are indicated.

#### *Shigella*

Tacket et al. (33) demonstrated the efficacy of a hyperimmune bovine colostrum against *Shigella flexneri* 2a in preventing infection in challenged volunteers. The duration of shedding of the organism was also decreased in the treatment group.

#### NONSPECIFIC DIARRHEA

In 1973, Fernandez et al. (16) reported the positive benefits of treating children with prolonged infantile diarrhea with lyophilized bovine colostrum.

A study reported from Prague (61) suggested that partly purified bovine colostrum with high volumes of antibodies against certain strains of enteropathogenic *E. coli* was beneficial in treating preterm and full-term infants hospitalized with diarrhea.

Stephan et al. (28) reported that 22 of 33 treatment cycles in 29 patients with AIDS receiving 10 g/day of bovine colostrum immunoglobulin from non-immunized cows for 10 days produced a significant decrease in nonspecific diarrhea. In a similar open and uncontrolled study, Rump et al. (32), using the same preparation in patients with AIDS with chronic diarrhea, found a significant clinical benefit, free of side effects, for the duration of therapy.

#### CONCLUSIONS

The results from the use of colostrum-derived bovine immunoglobulins in humans suggest that these immunoglobulins may have an important place in the prevention of many diarrheal disease. They have already been shown to be of value in preventing hospital-acquired symptomatic rotavirus infection (5, 6) and also in causing a reduction in rotavirus diarrhea in the community (8). *Clostridium difficile*, another important cause of hospital-acquired infection, particularly in adults and AIDS-infected patients, is also a candidate for passive prevention, as suggested by recent animal studies (36). This passive approach would also seem to be potentially useful in daycare centers where rotavirus, *Campylobacter*, *Shigella*, and *Cryptosporidium* are important pathogens (62).

The immunosuppressed patient, whether immunosuppressed due to a congenital defect or secondary to chemotherapy or infections such as AIDS, is particularly susceptible to a number of life-threatening enteric infections. Often no other therapy is available, and hyperimmune bovine colostrum has shown promise, particularly against *Cryptosporidium* (23,27,29,32) and rotavirus (9,10).

Another important possibility for the passive approach is in the prevention of travelers' diarrhea, which affects about half of all travelers to developing countries (63). Hyperimmune bovine immunoglobulin has been shown to be safe and effective in prevention of the major causes of travelers' diarrhea, including enterotoxigenic *E. coli* (25), *Shigella* (33), rotavirus (5-10,18,20), and *Campylobacter* (35). The current use of antibiotic prophylaxis, while effective, has side effects and is already leading to the development of significant drug resistance (64).

The possibility of the development of allergic reactions to bovine immunoglobulin, which has the potential to limit its usefulness, has been raised (64), and studies were carried out in known milk- and/or egg-allergic children. However, studies to date have not reported allergic reactions or any other adverse reaction. In several studies, children known to be allergic to cow milk were excluded (5,6), which would seem to be a sensible approach at present. The possibility of later development of atopic disease in young infants fed bovine immunoglobulin requires further study.

Hyperimmune bovine colostrum or immunoglobulin is relatively cheap and easy to produce, is straightforward to use, appears to be safe except possibly in allergic children, and may provide a simple approach to the prevention and possibly also treatment of many enteric pathogens. These facts are particularly important, since active vaccines for many pathogens seem to be a long way off. Even when active vaccines do become available, the use of passive protection will still be important in young infants and in children and adults who are unable to mount their own immune response. Active immunization of all children is difficult to achieve with current immunization programs, and passive protection in settings where children are at risk for cross-infection may remain beneficial.

## REFERENCES

- Kapikian AZ, Flores J, Hoshino Y, et al. Rotavirus: the major etiologic agent of severe infantile diarrhea may be controllable by a 'Jennerian' approach to vaccination. *J Infect Dis* 1986;153:815-22.
- Levine MM, Losonsky G, Herrington D, et al. Pediatric diarrhea: the challenge of prevention. *Pediatr Infect Dis J* 1986;5(suppl):529-43.
- Levine MM. Vaccines and milk immunoglobulin concentrates for prevention of infectious diarrhea. *J Pediatr* 1991; 118:S129-36.
- Ringenbergs M, Davidson GP, Spence J, Morris S. Prospective study of nosocomial rotavirus infection in a pediatric hospital. *Aust Paediatr J* 1989;25:156-60.
- Davidson GP, Whyte PBD, Daniels E, et al. Passive immunization of children with bovine colostrum containing antibodies to human rotavirus. *Lancet* 1989;2:709-12.
- Davidson GP, Tam J, Kirubakaran C. Passive protection against symptomatic hospital acquired rotavirus infection in India and Hong Kong. *J Pediatr Gastroenterol Nutr* 1994; 19:351.
- Barnes GL, Doyle LW, Hewson PH, et al. A randomized trial of oral gammaglobulin in low-birth-weight infants infected with rotavirus. *Lancet* 1982;1:1371-3.
- Turner RB, Kelsey DK. Passive immunization for prevention of rotavirus illness in healthy infants. *Pediatr Infect Dis J* 1993;12:718-72.
- Saulsbury FT, Winkelstein JA, Yolken RH. Chronic rotavirus infection in immunodeficiency. *J Pediatr* 1980;97:61-5.
- Losonsky G, Johnson JP, Winkelstein JA, Yolken RH. Oral administration of human serum immunoglobulin in immunodeficient patients with viral gastroenteritis. A pharmacokinetic and functional analysis. *J Clin Invest* 1985;76:2362-7.
- Offit PA, Clark HF. Maternal antibody-mediated protection against gastroenteritis due to rotavirus in newborn mice is dependent on both serotype and titer of antibody. *J Infect Dis* 1985;152:1152-8.
- Pickering LK, Ruiz-Palacios G. Antibodies in milk directed against specific enteric pathogens. In: Hamosh F, Goldman A, eds. *Human lactation: II*. New York: Raven Press, 1990: 499-506.
- Ringenbergs M, Albert MJ, Davidson GP, Goldworthy W, Haslam R. Serotype-specific antibodies to rotavirus in human colostrum and breast milk. *J Infect Dis* 1988;158:477-80.
- Glass RI, Stoll BJ, Wyatt RG, Hoshino Y, Banu H, Kapikian AZ. Observations questioning the protective role for breast feeding in severe rotavirus diarrhea. *Acta Paediatr Scand* 1986;75:713-18.
- Gurwith M, Wenman W, Hinde D, Feltham S, Greenberg H. A prospective study of rotavirus infection in infants and young children. *J Infect Dis* 1981;144:218-24.
- Fernandez LB, Averbach J, Ledesma de Paolo MI, Delle-done ME, Gonzalez E. Lyophilized bovine colostrum in the treatment of prolonged infantile diarrhea. *Am J Clin Nutr* 1973;26:383-4.
- Mietens C, Keinhorst H, Hilpert H, Gerber H, Amster H, Pahud JJ. Treatment of infantile *E. coli* gastroenteritis with specific bovine anti-*E. coli* milk immunoglobulins. *Eur J Pediatr* 1979;32:239-52.
- Ebina T, Sato A, Umezaki K, et al. Prevention of rotavirus infection by oral administration of cow colostrum containing anti-human rotavirus antibody. *Med Microbiol Immunol* 1985;174:177-85.
- Tzipori S, Robertson D, Chapman C. Remission of diarrhea due to cryptosporidiosis in an immunodeficient child treated with hyperimmune bovine colostrum. *Br Med J* 1986;293: 1276-7.
- Brussow H, Hilpert H, Walther I, Sidoti J, Mietens C, Bachmann P. Bovine milk immunoglobulins for passive immunity to infantile rotavirus gastroenteritis. *J Clin Microbiol* 1987; 25:982-6.
- Hilpert H, Brussow H, Mietens C, Sidoti J, Lemer L, Werchau H. Use of bovine milk concentrate containing antibody to rotavirus to treat rotavirus gastroenteritis in infants. *J Infect Dis* 1987;156:158-66.
- Saxon A, Weinstein W. Oral administration of bovine colostrum anticryptosporidia antibody fails to alter the course of human cryptosporidiosis. *J Parasitol* 1987;73:413-5.
- Tzipori S, Robertson D, Cooper DA, White L. Chronic cryptosporidial diarrhea and hyperimmune cow colostrum. *Lancet* 1987;2:344-5.
- McClead RE, Butler T, Rabbani GH. Orally administered bovine colostrum anti-cholera toxin antibodies: results of two clinical trials. *Am J Med* 1988;85:811-6.
- Tacket C, Losonsky G, Link H, et al. Protection by milk immunoglobulin concentrate against oral challenge with enterotoxigenic *Escherichia coli*. *N Engl J Med* 1988;318: 1240-3.
- Heaton P. Cryptosporidiosis and acute leukemia. *Arch Dis Child* 1990;65:813-4.
- Nord J, Ma P, Dijksh D, Tzipori S, Tacket CO. Treatment with bovine hyperimmune colostrum of cryptosporidial diarrhea in AIDS patients. *AIDS* 1990;4:581-4.
- Stephan W, Dichtelmüller H, Lissner R. Antibodies from colostrum in oral immunotherapy. *J Clin Chem Clin Biochem* 1990;28:19-23.

29. Ungar BLP, Ward DJ, Fayer R, Quinn CA. Cessation of cryptosporidium associated diarrhea in an acquired immunodeficiency syndrome patient after treatment with hyperimmune bovine colostrum. *Gastroenterology* 1990;98:486-9.
30. Brunser O, Espinoza J, Figueroa G, et al. Field trial of an infant formula containing anti-rotavirus and anti-*Escherichia coli* milk antibodies from hyperimmunized cows. *J Pediatr Gastroenterol Nutr* 1992;15:63-72.
31. Ebina T, Ohta M, Kanamaru Y, Yamamoto-Osumi Y, Baba K. Passive immunizations of suckling mice and infants with bovine colostrum containing antibodies to human rotavirus. *J Med Virol* 1992;38:117-23.
32. Rump JA, Arndt R, Arnold A, et al. Treatment of diarrhea in human immunodeficiency virus-infected patients with immunoglobulins from bovine colostrum. *Clin Invest* 1992;70:588-94.
33. Tacket CO, Binion SB, Bostwick E, Losonsky G, Roy MJ, Edelman R. Efficacy of bovine milk immunoglobulin concentrate in preventing illness after *Shigella flexneri* challenge. *Am J Trop Med Hyg* 1992;47:276-83.
34. Williams AC. Cryptosporidiosis in a child with leukemia. *Aust J Med Sci* 1992;13:176-9.
35. Husu J, Syaola E-L, Ahola-Luttilla H, Kalsta H, Sivela S, Kosunen TU. Production of hyperimmune bovine colostrum against *Campylobacter jejuni*. *J Appl Bacteriol* 1993;74:564-9.
36. Lyerly DM, Bostwick EF, Binion SB, Wilkins TD. Passive immunization of hamster against disease caused by *Clostridium difficile* by use of bovine immunoglobulin G concentrate. *Infect Immunol* 1991;59:2215-8.
37. Fayer R, Perryman LE, Riggs MW. Hyperimmune bovine colostrum neutralizes *Cryptosporidium* sporozoites and protects mice against oocyst challenge. *J Parasitol* 1989;75:151-3.
38. Arrowood MJ, Mead JR, Mahrt JL, Sterling CR. Effects of immune colostrum and orally administered antiparasite monoclonal antibodies on the outcome of *Cryptosporidium parvum* infections in neonatal mice. *Infect Immunol* 1989;57:2283-8.
39. Snodgrass DR, Wells PW. Rotavirus infection in lambs: studies on passive protection. *Arch Virol* 1976;52:201-5.
40. Saif LJ, Redman DR, Smith KL, Theil KW. Passive immunity to bovine rotavirus in newborn calves fed colostrum supplements from immunized or non-immunized cows. *Infect Immunol* 1983;41:1118-31.
41. Saif LJ. Passive immunity to coronavirus and rotavirus infection in swine and cattle: enhancement by maternal vaccination. In: Tzipori S, ed. *Infectious diarrhea in the young. Strategies for control in humans and animals*. Amsterdam: Excerpta Medica, 1985:456-67.
42. Lecce JG, Leary HL, Clare DA, Batema RP. Protection of agammaglobulinemic piglets from porcine rotavirus infection by antibody against simian rotavirus SA-11. *J Clin Microbiol* 1991;29:1382-6.
43. Schaller JP, Saif LJ, Cordle CT, Candler E Jr, Winship TR, Smith KL. Prevention of human rotavirus-induced diarrhea in gnotobiotic piglets using bovine antibody. *J Infect Dis* 1992;165:623-30.
44. Yoshiyama Y, Brown WR. Specific antibodies to cholera toxin in rabbit milk are protective against *Vibrio cholerae*-induced intestinal secretion. *Immunology* 1987;61:543-7.
45. Boesman-Finkelstein M, Walton NE, Finkelstein RA. Bovine lactogenic immunity against cholera toxin-related enterotoxins and *Vibrio cholerae* outer membranes. *Infect Immunol* 1989;57:1227-34.
46. Zinkernagel RM, Hilpert H, Gerber H. The digestion of colostrum bovine immunoglobulins in infants. *Experientia* 1972;28:741.
47. Petschow BW, Talbot RD. Reduction in virus-neutralizing activity of a bovine colostrum immunoglobulin concentrate by gastric acid and digestive enzymes. *J Pediatr Gastroenterol Nutr* 1994;19:228-35.
48. Pickering L. Post-partum maternal immunization with RRV and RRV tetraivalent. Presented at Fourth NIH Rotavirus Vaccine Workshop, Bethesda, MD, U.S.A., November 5-6, 1990.
49. Brussow H, Walther J, Fryder V, Sidoti J, Bruttin A. Cross-neutralization antibodies induced by a single serotype vaccination of cows with rotavirus. *J Gen Virol* 1988;69:1647-58.
50. Snodgrass DR, Ojeh CK, Campbell I, Herring AJ. Bovine rotavirus serotypes and their significance for immunization. *J Clin Microbiol* 1984;20:342-6.
51. Coulson BS, Tursi JM, McAdam WJ, Bishop RF. Derivation of neutralizing monoclonal antibodies to human rotaviruses and evidence that an immunodominant neutralization site is shared between serotypes 1 and 3. *Virology* 1986;154:302-12.
52. Green KY, Taniguchi K, Macow ER, Kapikian AZ. Homotypic and heterotypic epitope-specific antibody responses in adult and infant rotavirus vaccines: Implications for vaccine development. *J Infect Dis* 1990;161:667-79.
53. Ebina T, Tsukada K, Umezaki K, et al. Gastroenteritis in suckling mice caused by human rotavirus can be prevented by egg yolk immunoglobulin (IgY) and treated with a protein-bound polysaccharide preparation (PSK). *Microbiol Immunol* 1990;34:617-29.
54. Yolken RH, Leister F, Wee S-B, Miskuff R, Vonderfecht S. Antibodies to rotaviruses in chicken's eggs: a potential source of antiviral immunoglobulin suitable for human consumption. *Pediatrics* 1988;81:291-5.
55. Yolken RH, Peterson JA, Vonderfecht SL, Fouts ET, Midthun K, Newburg DS. Human milk mucin inhibits rotavirus replication and prevents experimental gastroenteritis. *J Clin Invest* 1992;90:1984-91.
56. Yolken RH, Ojeh C, Khatri IA, Sajjan LI, Forstner JF. Intestinal mucins inhibit rotavirus replication in an oligosaccharide-dependant manner. *J Infect Dis* 1994;169:1002-6.
57. Saavedra JM, Bauman NA, Oung I, Perman JA, Yolken RH. Feeding of Bifidobacterium bifidum and Streptococcus thermophilus to infants in hospital for prevention of diarrhea and shedding of rotavirus. *Lancet* 1994;344:1046-9.
58. Mitra AK, Mahalanabis D, Ashrof H, Unicomb L, Eeckels R, Tzipori S. Hyperimmune cow colostrum reduces diarrhea due to rotavirus: a double-blind controlled clinical trial. *Acta Paediatr* 1995;84:996-1001.
59. Guarino A, Canani RB, Russo S, et al. Oral immunoglobulins for treatment of acute rotaviral gastroenteritis. *Pediatrics* 1994;93:12-6.
60. Guarino A, Guandalini S, Albano F, Mascia A, deRitis G, Rubino A. Enteral immunoglobulins for treatment of protracted rotaviral diarrhea. *Pediatr Infect Dis J* 1991;10:612-4.
61. Lodinova-Zadnikova R, Korych B, Bartakova Z. Treatment of gastrointestinal infections in infants by oral administration of colostrum antibodies. *Die Nahrung* 1987;31:465-7.
62. Pickering LK, Bartlett AV, Woodward WE. Acute infectious diarrhea among children in day care: epidemiology and control. *Rev Infect Dis* 1986;8:439-47.
63. Consensus Development Conference statement. *Rev Infect Dis* 1986;8:5227-33.
64. Murray BE. Resistance of *Shigella*, *Salmonella*, and other selected enteric pathogens to antimicrobial agents. *Rev Infect Dis* 1986;8:5172-81.
65. Bernhisel-Broadbent J, Yolken RH, Sampson HA. Allergenicity of orally administered immunoglobulin preparations in food-allergic children. *Paediatrics* 1991;87:208-14.